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# Gene expression regulation and lineage evolution: the North and South tale of the hybrid polyploid *Squalius alburnoides* complex

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The evolution of hybrid polyploid vertebrates, their viability and their perpetuation over evolutionary time have always been questions of great interest. However, little is known about the impact of hybridization and polyploidization on the regulatory networks that guarantee the appropriate quantitative and qualitative gene expression programme. The *Squalius alburnoides* complex of hybrid fish is an attractive system to address these questions, as it includes a wide variety of diploid and polyploid forms, and intricate systems of genetic exchange. Through the study of genome-specific allele expression of seven housekeeping and tissue-specific genes, we found that a gene copy silencing mechanism of dosage compensation exists throughout the distribution range of the complex. Here we show that the allele-specific patterns of silencing vary within the complex, according to the geographical origin and the type of genome involved in the hybridization process. In southern populations, triploids of *S. alburnoides* show an overall tendency for silencing the allele from the minority genome, while northern population polyploids exhibit preferential biallelic gene expression patterns, irrespective of genomic composition. The present findings further suggest that gene copy silencing and variable expression of specific allele combinations may be important processes in vertebrate polyploid evolution.

**Keywords:** allopolyploid; gene expression; allele silencing; hybrid lineage evolution

## 1. INTRODUCTION

Polyploidy has been proposed as an important driving force of evolution, and the success of polyploid lineages confirms it as a successful evolutionary transition and a potentially relevant factor in evolutionary diversification (Otto & Whitton 2000). Ploidy rise brought upon by a hybridization event creates an epigenetic instability state that can only be overcome if regulation mechanisms, which contribute to gene copy perpetuation and heterosis, outrun disadvantages and potentiate species adaptation and viability. Therefore, it is important to investigate the functional basis of how polyploids overcome an initial period of instability, and establish processes that allow evolutionary flexibility and efficient competition with their diploid counterparts.

Hybrid polyploidy success has been documented in numerous plant species (Otto & Whitton 2000) and also in animals (Dowling & Secor 1997), but the regulatory changes that contribute to genome stabilization and regulation in the presence of distinct chromosome sets are still elusive. Successful polyploid vertebrates including

*Rana esculenta* (Hotz *et al.* 1999) and the *Bufo viridis* complex (Stöck *et al.* 2002), and asexual hybrid lineages such as the Amazon molly, *Poecilia formosa* (reviewed in Lampert & Scharl 2008), have been extensively studied regarding different molecular and reproductive aspects that contribute to their viability (Lamatsch & Stöck 2009). In all these systems, several aspects (such as the relevance of heterosis and the role of gene duplicates in lineage-specific evolution) have been comprehensively addressed (Comai 2005; Pignatta & Comai 2009), but the impact of polyploidization on the mechanisms regulating gene expression has not yet been clarified. Several deviations in the expected gene expression patterns have been reported in polyploid plants (Auger *et al.* 2005), including organ-specific silencing (Adams *et al.* 2003), epigenetic regulation of duplicates (Comai *et al.* 2000; Shaked *et al.* 2001), parent-of-origin-specific control of gene expression (Alleman & Doctor 2000) and a still unclear effect designated as ‘odd ploidy response’ (Guo *et al.* 1996). In vertebrates, little has been done to understand the effects of ploidy rise on gene regulation and their impact on the evolutionary potential of populations. The *Squalius alburnoides* complex was the first system in which dosage compensation by gene copy silencing was reported in a polyploid vertebrate context (Pala *et al.* 2008).

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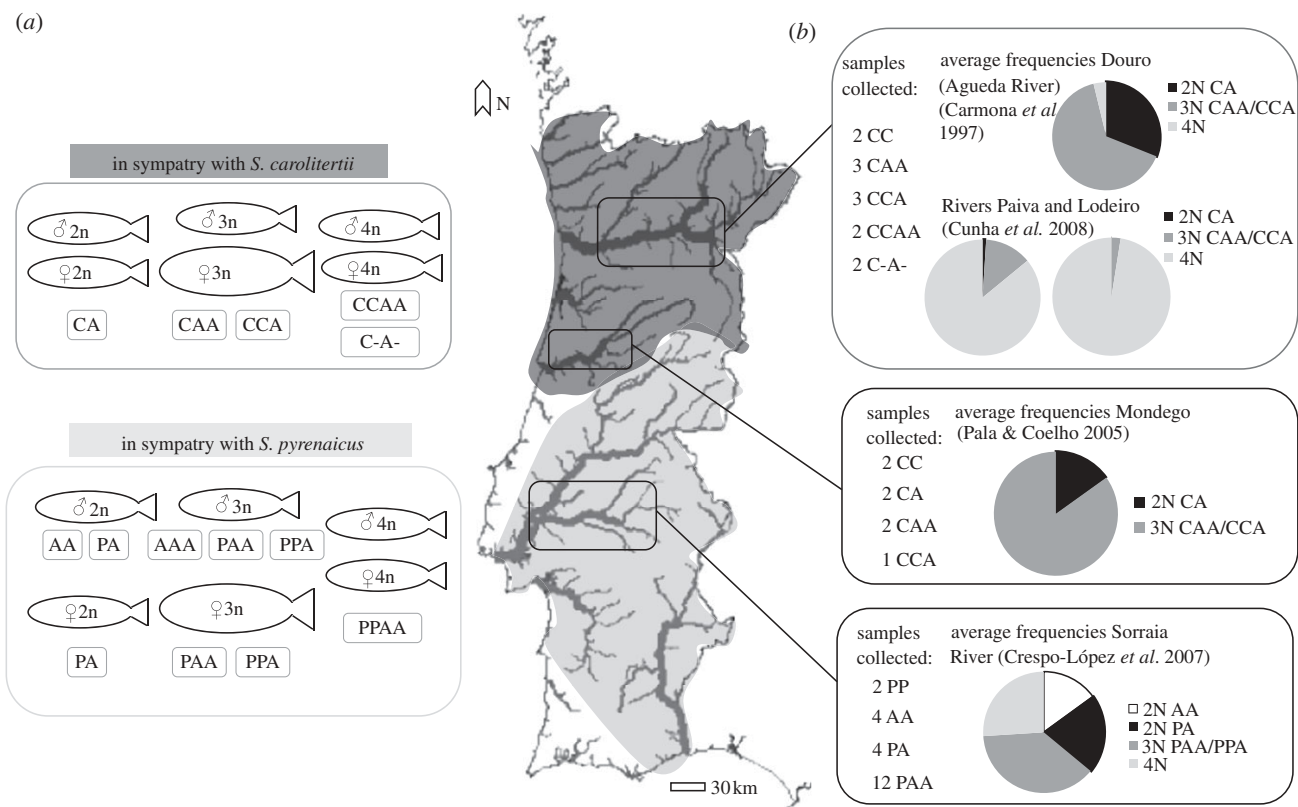


Figure 1. Distribution of *S. alburnoides* in the areas of sympatry with *S. carolitertii* (dark grey) and *S. pyrenaicus* (light grey). (a) The global composition of the populations of the complex in the two areas of sympatry are indicated, although local abundances can vary. (b) Samples collected in each location and reported frequencies of diploid (2N), triploid (3N) and tetraploid (4N) forms in Douro, Mondego and Tejo River basins.

This complex of hybrid fish presents several unique features that make it an ideal system to address the question of gene expression regulation and its impact on the persistence of hybrid lineages over evolutionary time (reviewed in Alves *et al.* 2001). It is an allopolyploid complex, resulting from interspecific hybridization between two Iberian cyprinid species: *Squalius pyrenaicus* as the maternal ancestor (contributing with the so-called P genome), and a still undetermined species (closely related to *Anaocypris hispanica*) as the paternal ancestor (A genome; reviewed in Alves *et al.* 2001). Presently, the complex is widely distributed in the Iberian Peninsula, where it occurs in sympatry with two species of the genus *Squalius*: *S. pyrenaicus* (P genome) in southern basins and *S. carolitertii* (C genome) in northern basins (figure 1a). These species form a reproductive complex with the hybrids, leading to diploid and polyploid offspring (figure S1, electronic supplementary material). The two bisexual species act as sources of new genetic material, and contribute to the maintenance of gene flow and the continuous cycling of genomes between forms (reviewed in Alves *et al.* 2001). The complex is composed of animals with different ploidy degrees and genomic constitutions, including diploids (PA, CA), triploids (PAA, PPA, CAA and CCA) and tetraploids, with C or P genome inclusion according to the geographical location (figure 1a). All forms are apparently fertile and interact through diverse reproductive modes (figure S1, electronic supplementary material) that include rare gynogenesis and processes of hybridogenesis (Carmona *et al.* 1997; Alves *et al.* 2001; Pala & Coelho 2005). The escape from

strictly asexual reproduction and the complexity of the genetic exchange routes definitively contribute to the success of *S. alburnoides*. In southern populations, an additional form designated as ‘nuclear non-hybrid’ (AA; carrying mtDNA of *S. pyrenaicus*) occurs and actively contributes to the maintenance of the genetic diversity of the complex (Alves *et al.* 1999).

We used the *S. alburnoides* system to approach the question of gene expression regulation and evolution of polyploid taxa. As an allopolyploid, the *S. alburnoides* complex offers the advantage of allowing the distinction between the different genome-specific gene copies and how they contribute to overall expression. The presence of lineages within the complex, established differentially in time, including different forms and showing distinct dynamics of genetic exchange and different evolutionary potential, further allows us to explore the question of whether the mechanisms of gene silencing observed in triploids of the southern populations of *S. alburnoides* (Pala *et al.* 2008) would also be present in other independent populations and in different genomic compositions.

We have studied the expression pattern of a total of seven widely expressed and tissue-restricted genes in different organs of diploid, triploid and tetraploid forms from southern and northern populations of the complex. We have found differential expression patterns of genome-specific alleles according to geographical location and have attempted to integrate the observed disparities with the distinct genomic and evolutionary features of the different populations of *S. alburnoides*. We put forward

the hypothesis that the type of genomes brought together upon the hybridization process might be an important factor in the establishment of specific mechanisms of gene expression regulation in vertebrate allopolyploids.

## 2. MATERIAL AND METHODS

### (a) *Samples, DNA and RNA extraction, and genotyping*

Samples of *S. alburnoides* and of the sympatric *Squalius* species were collected from three locations corresponding to the northern and southern distribution ranges of the complex (figure 1*b*). Individuals were sacrificed with an overdose of the anaesthetic MS222. Organs and fin clips were collected for RNA and DNA extraction, which were performed as described in Pala *et al.* (2008). Blood samples were drawn from the caudal vein, stabilized in buffer (40 mM citric acid trisodium salt, 0.25 M sucrose and 5% dimethyl sulphoxide) and immediately frozen at  $-80^{\circ}\text{C}$ . The determination of genotype identity of individual samples was performed by a conjoined approach of flow-cytometry measurements (Dawley & Goddard 1988) and the analysis of microsatellite variation.

Genotypes were determined by cross-species amplification of three microsatellite loci (LCO3, LCO4 and LCO5) using the Multiplex PCR Kit (Qiagen) and identification of alleles specific to the respective genomes (P, C and A), as previously described (Pala & Coelho 2005; Crespo-López *et al.* 2007; Cunha *et al.* 2008). In the cases for which it was not possible to identify the three or four distinct alleles in triploids and tetraploids, an additional locus (LCO1) was used (table S1, electronic supplementary material). Amplification products were analysed with an automated sequencer (ABI 310 Genetic Analyser).

### (b) *Sequence determination and genome expression*

Sequences of known teleost orthologues of a total of seven genes—ubiquitously expressed ( *$\beta$ -actin*, *rpl8*, *ef1a* and *gapdh*), gonad-specific (*amh*, *dmrt1*) and eye-specific (*rhodopsin*)—were used as templates for the design of gene-specific primers (table S2, electronic supplementary material). Primers were initially tested on cDNA samples of *S. pyrenaicus* (PP), *S. carolitertii* (CC) and *S. alburnoides* (AA). Amplifications were performed according to the following PCR conditions: pre-heating at  $94^{\circ}\text{C}$  for 2 min 30 s, 35 cycles at  $94^{\circ}\text{C}$  for 45 s,  $52^{\circ}\text{C}$  (*amh*, *dmrt1*)/ $55^{\circ}\text{C}$  (*rhodopsin*,  *$\beta$ -actin*, *rpl8*, *ef1a* and *gapdh*) for 40 s and  $72^{\circ}\text{C}$  for 1 min 15 s, and a final extension at  $72^{\circ}\text{C}$  for 10 min. Polymorphic sites for the three genomes (P, C and A) were identified for the seven genes by sequence alignment using SEQUENCHER v. 4.0 (Gene Codes Corporation). In hybrid samples, the presence of cDNAs derived from single genome copies or from both genomes was determined through sequence comparison and based on the identified polymorphic sites between genomes (P, C and A). Genome control sequences, representing the P, C and A genomes and obtained from *S. pyrenaicus*, *S. carolitertii* and nuclear non-hybrid *S. alburnoides*, were also analysed. Two to four forward and reverse sequences of each gene were obtained per individual/per organ, using independently synthesized cDNA samples and PCR amplifications. PCR amplifications for replicate reactions were also performed independently, following the procedures described in Pala *et al.* (2008).

### (c) *Quantitative real-time PCR analysis*

Primers and specific TaqMan probes for  *$\beta$ -actin*, *rpl8*, *gapdh* (Pala *et al.* 2008) and *ef1a* (EFreal-F5'-CCGTC TGCCACTTCAGGATG-3'; EFreal-R5'-CATACCAGGC TT GAGGACACC-3'; EF/6VIC 5'-TCCACACGACCC ACGGGCACAGT-3') were used for amplification of muscle, liver and gonad samples of representative specimens of the CAA and CCA triploid (3–5) and CCAA tetraploid (1–2) forms from the Douro and Mondego River basins. Reactions and analysis were conducted as described in Pala *et al.* (2008).

## 3. RESULTS

### (a) *Gene expression patterns differ according to organ and geographical location*

The sequence analysis of genome-specific polymorphisms for the different genes in the various organs revealed contrasting results between southern and northern populations of *S. alburnoides* (table 1).

In the sample from the southern population (Tejo Basin), the four housekeeping genes ( *$\beta$ -actin*, *rpl8*, *ef1a* and *gapdh*) showed differential expression patterns according to organ. In muscle and liver, diploid PA individuals showed the expected simultaneous expression of A and P alleles (referred from now on as biallelic expression). Triploids, on the other hand, exhibited a variety of expression profiles: for  *$\beta$ -actin* and *rpl8* most muscle and liver samples exhibited expression exclusively from the A genome, although biallelic expression was also observed. For *ef1a*, two triploid samples in muscle and one in liver showed biallelic expression while in the remaining one only A genome transcripts were identified. For *gapdh*, biallelic expression of P and A alleles was identified in all triploid muscle samples, while in liver samples all genes exhibited a predominantly monoallelic expression of A genome transcripts, with a small number of individuals exhibiting biallelic expression. The most paradigmatic example of this was the  *$\beta$ -actin* gene, of which transcripts in triploids corresponded exclusively to the A genome.

In individuals from northern populations (Mondego and Douro Basins), in liver and muscle samples, biallelic expression was observed with no exceptions for all gene/organ combinations irrespective of ploidy level and genomic constitution, including CA, CCA, CAA and CCAA *S. alburnoides*. The same was true for  *$\beta$ -actin* and *rpl8* transcripts in eye samples of triploid (CCA and CAA) and tetraploid individuals: both C and A alleles were expressed (figure S2, electronic supplementary material). A tendency towards a preferential biallelic expression was also observed in eye samples of southern triploid (PAA) specimens: four out of seven samples showed expression of P and A alleles of the  *$\beta$ -actin* gene, and *rhodopsin* transcripts resulted from biallelic expression in all samples analysed.

Brain samples from triploid and tetraploid individuals from Mondego and Douro showed an exception to the overall tendency of biallelic expression of C and A genomes in these basins: although C and A  *$\beta$ -actin* expression was observed in all samples, the *gapdh* gene showed monoallelic A-genome-exclusive expression in CAA individuals (figure S2, electronic supplementary material).



Table 1. Relative frequency of allele-specific transcripts of P, C and A genomes in different organs of individuals from southern (Tejo) and northern (Mondego and Douro) populations of the complex.

<i>Tejo (south)</i>				muscle			liver			gonad		
ploidy	species	genotype		P	A	PA	P	A	PA	P	A	PA
2n	<i>S. pyrenaicus</i>	PP	across all genes	1	0	0	1	0	0	1	0	0
	<i>S. alburnoides</i>	AA		0	1	0	0	1	0	0	1	0
	<i>S. alburnoides</i>	PA		0	0	1	0	0	1	0	0	1 <sup>a</sup>
3n	<i>S. alburnoides</i>	PAA	<i>β-actin</i>	0	0.67	0.33	0	1	0	0	1	0
			<i>rpl8</i>	0	0.67	0.33	0	0.75	0.25	0	1	0
			<i>ef1a</i>	0	0.33	0.67	0	0.83	0.17	0	1	0
			<i>gapdh</i>	0	0	1	0	0.64	0.36	0	1	0
			<i>amh</i>	—	—	—	—	—	—	0	0	1
			<i>dmrt1</i>	—	—	—	—	—	—	0	0.75	0.25
<i>Mondego/Douro (north)</i>												
ploidy	species	genotype		C	A	CA	C	A	CA	C	A	CA
2n	<i>S. carolitertii</i>	CC	across all genes	1	0	0	1	0	0	1	0	0
	<i>S. alburnoides</i>	CA		0	0	1	0	0	1	0	0	1 <sup>a</sup>
3n	<i>S. alburnoides</i>	CAA	<i>β-actin</i>	0	0	1	0	0	1	0	1	0
			<i>rpl8</i>	0	0	1	0	0	1	0	1	0
			<i>ef1a</i>	0	0	1	0	0	1	0	1	0
			<i>gapdh</i>	0	0	1	0	0	1	0	1	0
			<i>amh</i>	—	—	—	—	—	—	0	0	1
			<i>dmrt1</i>	—	—	—	—	—	—	0	1	0
3n	<i>S. alburnoides</i>	CCA	across all genes	0	0	1	0	0	1	0	0	1
4n	<i>S. alburnoides</i>	CCAA		0	0	1	0	0	1	0	0	1
		C-A-		0	0	1	0	0	1	0	0	1

<sup>a</sup>Only *amh*, *dmrt1* and *β-actin* were analysed.

In adult gonads (table 1) of southern triploid individuals, all four housekeeping genes (*β-actin*, *rpl8*, *ef1a* and *gapdh*) showed exclusive expression of A genome alleles. The *amh* gene showed biallelic expression in triploid PAA, while *dmrt1* exhibited both monoallelic A expression (nine samples) and biallelic (three samples) genome expression. Diploid PA exhibited a biallelic expression pattern in all genes analysed (*amh*, *dmrt1* and *β-actin*). In northern samples, patterns were divergent. The *amh* gene was the only gene for which expression of both C and A alleles was observed in all diploid, triploid and tetraploid forms of *S. alburnoides*. The remaining five genes exhibited a distinctive expression profile between the two forms of triploids: while in CCA samples the observed expression pattern was biallelic (as in diploid and tetraploid samples from the same locations), in CAA individuals *dmrt1*, *β-actin*, *rpl8*, *ef1a* and *gapdh* expression was exclusively from the A genome. Exclusive expression of alleles from the minority genome was never observed in triploids.

#### (b) Quantitative expression

The relative expression ratios obtained from the comparison between average ct values of triploid and tetraploid samples and the diploid controls were always approximately 1 (figure 2), implying dosage compensation by regulation of gene expression to the diploid level in polyploids. Despite qualitative differences in allele representation, a similar effect of dosage compensation was observed in these populations compared with the one described in the southern Tejo basin (Pala *et al.* 2008).

The method was further validated by an independent quantitative real-time PCR analysis comparing definite cell numbers (I. Matos & M. M. Coelho 2009, unpublished data).

#### 4. DISCUSSION

In the present work, we have addressed the question of whether the mechanisms of gene silencing and regulation of transcripts to the diploid level, described in triploids of the southern populations of *S. alburnoides* (Pala *et al.* 2008), would also be present in other populations and in different genomic contexts. We have studied the expression patterns of housekeeping and tissue-specific genes in individuals of different genomic composition and ploidy degrees, from different locations within the distribution range of the *S. alburnoides* complex. Based on a representative sample of the forms that constitute the complex in different locations, we have attempted to understand the global effects of hybridization and ploidy rise on gene expression regulation in this system and evaluate their evolutionary implications.

The data obtained in the present work suggest a substantial difference in the genome-specific allele usage, depending on the geographical location of individuals. While a preferential expression of A genome and silencing of P genome alleles was observed in most triploids of southern populations of the complex, the vast majority of samples of the two northern river basins exhibited simultaneous expression of both C and A genome alleles, irrespective of ploidy level or genomic composition.

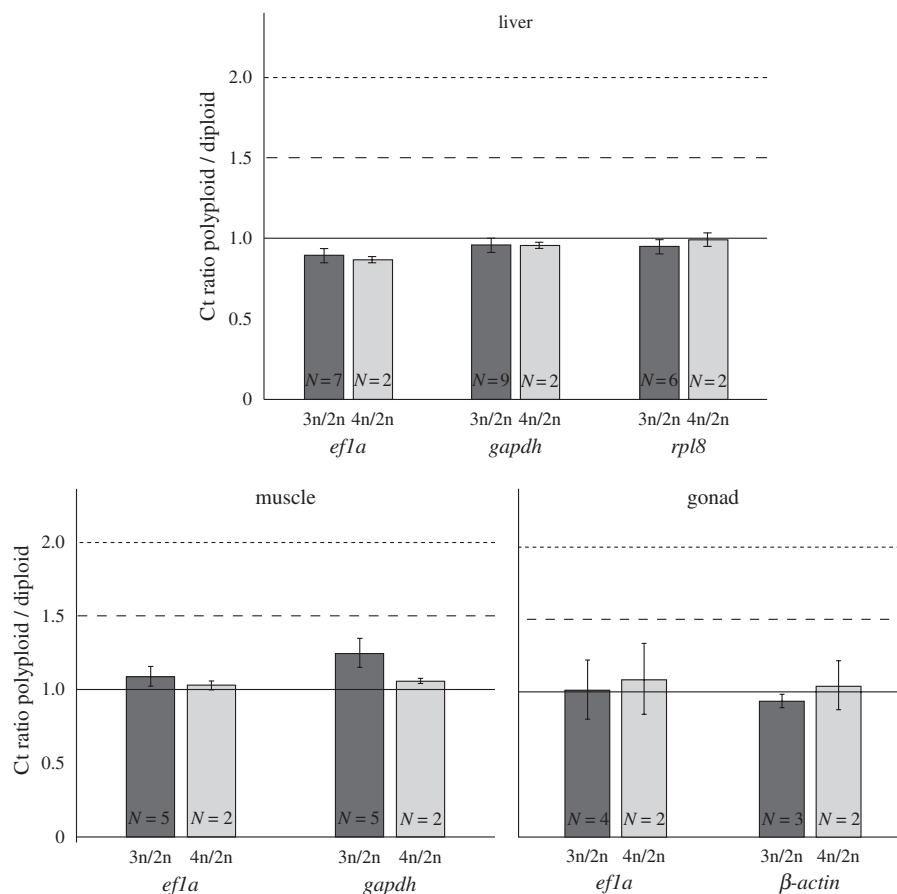


Figure 2. Relative expression (Ct) ratios between polyploid and diploid organ samples of individuals of the northern distribution of the complex, for the *ef1a*, *gapdh*, *rpl8* and *β-actin* genes. Expected ratio of 1.5 in the case of no silencing 3n/2n (dashed line), expected ratio of 2, in the case of no silencing 4n/2n (dotted line) and expected ratio of 1 in the case of silencing (solid line). Observed 3n to 2n ratio (dark grey columns), observed 4n to 2n ratio (light grey columns), number of triploid and tetraploid triplicates analysed for each ratio calculation (N).

#### (a) Gene expression and dosage compensation

In southern populations of the Tejo River basin, the problem of keeping balance of expression regulatory networks in an uneven-numbered genome is apparently overcome by allele silencing (Pala *et al.* 2008), similar to what has been observed in plant species following polyploidization (Adams *et al.* 2003, 2004). In plants, it has been proposed that adapted polyploids would avoid extinction by reducing gene redundancy and thus undergoing an evolutionary pathway leading to functional diploidization (Paterson *et al.* 2004; Wang *et al.* 2005). The need for correct gene balance in a polyploid context as a way to tolerate the presence of extra genome copies and to maintain stable aneuploids has been further demonstrated in plants (Birchler & Veita 2007) and discussed also as a possibility in vertebrates (Mable 2007). Recently, dosage effects on transcription levels have been revealed in salmon (Ching *et al.* 2010), and similar levels of gene expression have been reported in triploid and diploid individuals. In the *S. alburnoides* complex, the necessity for balanced gene expression is apparent in both southern (Pala *et al.* 2008) and northern populations, with higher ploidy forms exhibiting gene expression regulated to the diploid levels. The similar quantitative outcome, apparently ensured in both geographical locations, is nevertheless accompanied by a distinct usage of specific alleles. In plant allopolyploids, further support for the possibility of different genome-specific allele usage producing similar transcript levels comes from the study of genome-wide expression dominance. The expression

of one or the other genome was found to be dependent on the specific genomic combinations that were brought together in the hybrid, irrespective of the level of gene expression (Rapp *et al.* 2009).

Thus, the preferential monoallelic expression observed in the southern triploids of *S. alburnoides*, and the usage of both C and A allele copies in northern populations, are both in line with gene expression patterns observed in other polyploid taxa. Variable patterns of expression according to organ, silencing of genome specific alleles and non-additive gene expression have all been observed in plant polyploids (Adams *et al.* 2004; Auger *et al.* 2005; Paterson 2005; Salmon *et al.* 2005) and pointed out as consequences of the integration of divergent regulatory hierarchies.

Population differences in gene expression patterns of polyploids of the same species and ploidy level have also been reported in allopolyploid plants of the genus *Tragopodon* (Tate *et al.* 2006), and independent polyploidy events have been pointed out as a likely factor contributing to the observed variation. A similar effect apparently occurs in the *S. alburnoides* fish complex: patterns of gene expression diverge even within the same hybrid system, as shown by inequality of allele usage according to geographical origin and genome combination. Distinctive patterns of gene expression were also observed in gonads of the two triploid forms (CAA, CCA) in northern populations, contrasting with the overall identity in expression patterns in somatic organs. The patterns of

gene expression in CAA gonads were similar to the ones observed in southern PAA samples. This may be related to the type of reproduction and mechanism of gamete production of the different forms, namely the exclusion of the minority genome (Carmona *et al.* 1997; Alves *et al.* 1998; Pala & Coelho 2005). Conversely, the overall biallelic expression observed in CCA females is not so easy to integrate. Owing to their low abundance in most populations, no extensive information about the reproductive modes of CCA females has been gathered, and the present results might indicate that they do not follow the same process of hybridogenesis as other triploid females (Crespo-López *et al.* 2006).

### (b) Hybridization events and genome usage

The variation in allele expression dynamics observed here could be related to the difference in the origin and the timing of constitution of the three lineages of the complex (ranging from less than 0.7 to 0.01 Myr, according to Sousa-Santos *et al.* 2007). Regulation of gene expression can be variable in ancient and newly formed polyploids (Wang *et al.* 2004; Adams & Wendel 2005), and the disparity in the timing of the onset of hybridization events could be a factor to be taken into account.

An alternative possibility could be the difference in the composition of the genomes involved in the maintenance of the complex in each of the three basins. The C genome, introduced through the bisexual species *S. caroliertii* in northern populations, was a later addition to the complex, after initial hybridization events involving only *S. pyrenaicus* (Alves *et al.* 1997; Cunha *et al.* 2004). Genomic stress induced by two distinct genomes being brought together in the same cell nucleus often leads to different epigenetic modifications of homeologous (Riddle & Birchler 2003) and differential capacity of regulatory interactions (Comai 2000; Adams & Wendel 2004; Rapp *et al.* 2009). Both the P and C genome apparently have a good functional 'affinity' with the A genome, but it is possible that regulatory elements that have to interact with both heterologous genomes in hybrids respond differentially or have distinct interaction capacity.

Another important difference between the northern and southern populations of the complex, which has been pointed out as a key factor in the maintenance of its diversity and contributing actively to the evolutionary success of *S. alburnoides*, is the presence of nuclear non-hybrid males of AA genotype (Alves *et al.* 2002; Crespo-López *et al.* 2006, 2007). The preferential expression of A genome alleles in southern populations could be related to the establishment of this nuclear 'non-hybrid' lineage. A different preference for expressed alleles might have been established in northern populations in which the process of A allele recombination and shifting, mediated by the nuclear non-hybrid form, is not present.

The heterogeneity of the genomes brought together in plant allopolyploids has been proposed as a major factor underlying patterns of non-additive gene expression, as suggested by studies in different ploidy levels of cotton (Flagel *et al.* 2008) and *Senecio* (Hegarty *et al.* 2006). In these species, the effects of genome merger apparently supplant the impact of genome doubling. The differential patterns of gene expression according to genomic composition, reported here for the *S. alburnoides* complex, also

point towards a strong influence of the type of genomes involved in the hybridization events occurring in each geographical location. However, this might not be the sole explanation for the observed patterns. Dosage constraints and ploidy level might be additional key players in the regulation of gene expression in this system, as indicated by the similar behaviour of diploids of different geographical origins in terms of qualitative and quantitative gene expression, and the switch to differential patterns of genome specific allele usage in triploids of the northern and southern distribution of *S. alburnoides*.

The elucidation of the mechanism underlying dosage compensation and preferential allele usage, as well as the characterization of the factors that modulate differential gene expression, will hopefully contribute to a better understanding of how vertebrate polyploid genomes are regulated and how the different adopted strategies can influence their odds on the race for adaptability and evolutionary success.

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### REFERENCES

- Adams, K. L. & Wendel, J. F. 2004 Exploring the genomic mysteries of polyploidy in cotton. *Biol. J. Linn. Soc.* **82**, 573–582. (doi:10.1111/j.1095-8312.2004.00342.x)
- Adams, K. L. & Wendel, J. F. 2005 Polyploidy and genome evolution in plants. *Curr. Opin. Plant Biol.* **8**, 135–141. (doi:10.1016/j.pbi.2005.01.001)
- Adams, K. L., Cronn, R., Percifield, R. & Wendel, J. F. 2003 Genes duplicated by polyploidy show unequal contributions to the transcriptome and organ-specific reciprocal silencing. *Proc. Natl Acad. Sci. USA* **100**, 4649–4654. (doi:10.1073/pnas.0630618100)
- Adams, K. L., Percifield, R. & Wendel, J. F. 2004 Organ-specific silencing of duplicated genes in a newly synthesized cotton allotetraploid. *Genetics* **168**, 2217–2226. (doi:10.1534/genetics.104.033522)
- Alleman, M. & Doctor, J. 2000 Genomic imprinting in plants: observations and evolutionary implications. *Plant Mol. Biol.* **43**, 147–161. (doi:10.1023/A:1006419025155)
- Alves, M. J., Coelho, M. M., Collares-Pereira, M. J. & Dowling, T. E. 1997 Maternal ancestry of the *Rutilus alburnoides* complex (Teleostei, Cyprinidae) as determined by analysis of cytochrome *b* sequences. *Evolution* **51**, 1584–1592.
- Alves, M. J., Coelho, M. M. & Collares-Pereira, M. J. 1998 Diversity in the reproductive modes of females of the *Rutilus alburnoides* complex (Teleostei, Cyprinidae): a way to avoid the genetic constraints of uniparentalism. *Mol. Biol. Evol.* **15**, 1233–1242.
- Alves, M. J., Coelho, M. M., Próspero, M. I. & Collares-Pereira, M. J. 1999 Production of fertile unreduced sperm by hybrid males of the *Rutilus alburnoides* complex (Teleostei, Cyprinidae): an alternative route to genome tetraploidization in unisexuals. *Genetics* **151**, 277–283.
- Alves, M. J., Coelho, M. M. & Collares-Pereira, M. J. 2001 Evolution in action through hybridisation and polyploidy in an Iberian freshwater fish: a genetic review. *Genetica* **111**, 375–385. (doi:10.1023/A:1013783029921)

- Alves, M. J., Collares-Pereira, M. J., Dowling, T. E. & Coelho, M. M. 2002 The genetics of maintenance of an all-male lineage in the *Squalius alburnoides* complex. *J. Fish Biol.* **60**, 649–662. (doi:10.1111/j.1095-8649.2002.tb01691.x)
- Auger, D. L., Gray, A. D., Ream, T. S., Kato, A., Coe, E. H. & Birchler, J. A. 2005 Nonadditive gene expression in diploid and triploid hybrids of maize. *Genetics* **169**, 389–397. (doi:10.1534/genetics.104.032987)
- Birchler, J. A. & Veitia, R. A. 2007 The gene balance hypothesis: from classical genetics to modern genomics. *Plant Cell* **19**, 395–402. (doi:10.1105/tpc.106.049338)
- Carmona, J. A., Sanjurjo, O. I., Doadrio, I., Machordom, A. & Vrijenhoek, R. C. 1997 Hybridogenetic reproduction and maternal ancestry of polyploid Iberian fish: the *Tropidophoxinellus alburnoides* complex. *Genetics* **146**, 983–993.
- Ching, B., Jamieson, S., Heath, J. W., Heath, D. D. & Hubberstey, A. 2010 Transcriptional differences between triploid and diploid Chinook salmon (*Oncorhynchus tshawytscha*) during live *Vibrio anguillarum* challenge. *Heredity* **104**, 224–234. (doi:10.1038/hdy.2009)
- Comai, L. 2000 Genetic and epigenetic interactions in allopolyploid plants. *Plant Mol. Biol.* **43**, 387–399. (doi:10.1023/A:1006480722854)
- Comai, L. 2005 The advantages and disadvantages of being polyploid. *Nat. Rev. Genet.* **6**, 836–846. (doi:10.1038/nrg1711)
- Comai, L., Tyagi, A. P., Winter, K., Holmes-Davis, R., Reynolds, S. H., Stevens, Y. & Byers, B. 2000 Phenotypic instability and rapid gene silencing in newly formed *Arabidopsis* allotetraploids. *Plant Cell* **12**, 1551–1568. (doi:10.1105/tpc.12.9.1551)
- Crespo-López, M. E., Duarte, T., Dowling, T. & Coelho, M. M. 2006 Modes of reproduction of the hybridogenetic fish *Squalius alburnoides* in the Tejo and Guadiana rivers: an approach with microsatellites. *Zoology (Jena)* **109**, 277–286. (doi:10.1016/j.zool.2006.03.008)
- Crespo-López, M. E., Pala, I., Duarte, T., Dowling, T. E. & Coelho, M. M. 2007 Genetic structure of the diploid–polyploid fish *Squalius alburnoides* in southern Iberian basins Tejo and Guadiana, based on microsatellites. *J. Fish Biol.* **71**, 423–436. (doi:10.1111/j.1095-8649.2007.01688.x)
- Cunha, C., Coelho, M. M., Carmona, J. A. & Doadrio, I. 2004 Phylogeographical insights into the origins of the *Squalius alburnoides* complex via multiple hybridisation events. *Mol. Ecol.* **13**, 2807–2817. (doi:10.1111/j.1365-294X.2004.02283.x)
- Cunha, C., Doadrio, I. & Coelho, M. M. 2008 Speciation towards tetraploidization after intermediate processes of non-sexual reproduction. *Phil. Trans R. Soc. B* **363**, 2921–2929. (doi:10.1098/rstb.2008.0048)
- Dawley, R. M. & Goddard, K. A. 1988 Diploid–triploid mosaics among unisexual hybrids of the minnow *Phoxinus phoxinus*. *Evolution* **42**, 649–659.
- Dowling, T. E. & Secor, C. L. 1997 The role of hybridisation and introgression in the diversification of animals. *Annu. Rev. Ecol. Syst.* **28**, 593–620. (doi:10.1146/annurev.ecolsys.28.1.593)
- Flagel, L., Udall, J., Nettleton, D. & Wendel, J. 2008 Duplicate gene expression in allopolyploid *Gossypium* reveals two temporally distinct phases of expression evolution. *BMC Biol.* **6**, 16. (doi:10.1186/1741-7007-6-16)
- Guo, M., Davis, D. & Birchler, J. A. 1996 Dosage effects on gene expression in a maize ploidy series. *Genetics* **142**, 1349–1355.
- Hegarty, M. J., Barker, G. L., Wilson, I. D., Abbott, R. J., Edwards, K. J. & Hiscock, S. J. 2006 Transcriptome shock after interspecific hybridization in *Senecio* is ameliorated by genome duplication. *Curr. Biol.* **16**, 1652–1659. (doi:10.1016/j.cub.2006.06.071)
- Hotz, H., Semlitsch, R. D., Gutmann, E., Guex, G. D. & Beerli, P. 1999 Spontaneous heterosis in larval life-history traits of hemiclinal frog hybrids. *Proc. Natl Acad. Sci. USA* **96**, 2171–2176. (doi:10.1073/pnas.96.5.2171)
- Lamatsch, D. K. & Stöck, M. 2009 Sperm-dependent parthenogenesis and hybridogenesis in teleost fishes. In *Lost sex! The evolutionary biology of parthenogenesis* (eds I. Schön, K. Martens & P. Van Dijk), pp. 399–432. Berlin, Germany: Springer.
- Lampert, K. P. & Scharl, M. 2008 The origin and evolution of a unisexual hybrid: *Poecilia formosa*. *Phil. Trans. R. Soc. B* **363**, 2901–2909. (doi:10.1098/rstb.2008.0040)
- Mable, B. K. 2007 Sex in the postgenomic era. *Trends Ecol. Evol.* **22**, 559–561. (doi:10.1016/j.tree.2007.07.006)
- Otto, S. P. & Whitton, J. 2000 Polyploid incidence and evolution. *Annu. Rev. Genet.* **34**, 401–437. (doi:10.1146/annurev.genet.34.1.401)
- Pala, I. & Coelho, M. M. 2005 Contrasting views over a hybrid complex: between speciation and evolutionary ‘dead-end’. *Gene* **347**, 283–294. (doi:10.1016/j.gene.2004.12.010)
- Pala, I., Coelho, M. M. & Scharl, M. 2008 Dosage compensation by gene copy silencing in a triploid hybrid fish. *Curr. Biol.* **18**, 1344–1348. (doi:10.1016/j.cub.2008.07.096)
- Paterson, A. H. 2005 Polyploidy, evolutionary opportunity, and crop adaptation. *Genetica* **123**, 191–196. (doi:10.1007/s10709-003-2742-0)
- Paterson, A. H., Bowers, J. E. & Chapman, B. A. 2004 Ancient polyploidisation predating divergence of the cereals, and its consequences for comparative genomics. *Proc. Natl Acad. Sci. USA* **101**, 9903–9908. (doi:10.1073/pnas.0307901101)
- Pignatta, D. & Comai, L. 2009 Parental squabbles and genome expression: lessons from the polyploids. *J. Biol.* **8**, 43. (doi:10.1186/jbiol140)
- Rapp, R. A., Udall, J. A. & Wendel, J. F. 2009 Genomic expression dominance in allopolyploids. *BMC Biol.* **7**, 18. (doi:10.1186/1741-7007-7-18)
- Riddle, N. C. & Birchler, J. A. 2003 Effects of reunited diverged regulatory hierarchies in allopolyploids and species hybrids. *Trends Genet.* **19**, 597–600. (doi:10.1016/j.tig.2003.09.005)
- Salmon, A., Ainouche, M. L. & Wendel, J. F. 2005 Genetic and epigenetic consequences of recent hybridisation and polyploidy in *Spartina* (Poaceae). *Mol. Ecol.* **14**, 1163–1175. (doi:10.1111/j.1365-294X.2005.02488.x)
- Shaked, H., Kashkush, K., Ozkan, H., Feldman, M. & Levy, A. A. 2001 Sequence elimination and cytosine methylation are rapid and reproducible responses of the genome to wide hybridisation and allopolyploidy in wheat. *Plant Cell* **13**, 1749–1759. (doi:10.1105/tpc.13.8.1749)
- Sousa-Santos, C., Collares-Pereira, M. J. & Almada, V. 2007 Reading the history of a hybrid fish complex from its molecular record. *Mol. Phylogenet. Evol.* **45**, 981–996. (doi:10.1016/j.ympev.2007.05.011)
- Stöck, M. *et al.* 2002 A bisexually reproducing all-triploid vertebrate. *Nat. Genet.* **30**, 325–328. (doi:10.1038/ng839)
- Tate, J. A., Ni, Z., Scheen, A. C., Koh, J., Gilbert, C. A., Lefkowitz, D., Chen, Z. J., Soltis, P. S. & Soltis, D. E. 2006 Evolution and expression of homeologous loci in *Tragopogon miscellus* (Asteraceae), a recent and reciprocally formed allopolyploid. *Genetics* **173**, 1599–1611. (doi:10.1534/genetics.106.057646)
- Wang, J. *et al.* 2004 Stochastic and epigenetic changes of gene expression in *Arabidopsis* polyploids. *Genetics* **167**, 1961–1973. (doi:10.1534/genetics.104.027896)
- Wang, X., Shi, X., Hao, B., Ge, S. & Luo, J. 2005 Duplication and DNA segmental loss in the rice genome: implications for diploidisation. *New Phytol.* **165**, 937–946. (doi:10.1111/j.1469-8137.2004.01293.x)